

Therapeutical effects of PPAR agonists assessed by biomarker modulation

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Abstract

The metabolic syndrome is defined as the clustering of cardiovascular risk factors, such as glucose intolerance, hyperinsulinemia, dyslipidemia, coagulation disturbances and hypertension. Activators of the nuclear receptors peroxisome proliferator-activated receptors (PPARs) modulate several of the metabolic risk factors pre-disposing to atherosclerosis. Fibrates are hypolipidemic drugs operating through activation of PPAR α , whereas glitazones are insulin sensitizers activating PPARy. In addition, these drugs exert pleiotropic and anti-inflammatory actions. This review will focus on the different effects of fibrates and glitazones, as measured by biomarker modulation, on the development of atherosclerosis and cardiovascular disease.

Keywords: Nuclear receptors, fibrates, glitazones, metabolic syndrome

Introduction

The metabolic syndrome, which is characterized by the simultaneous presence of one or more metabolic disorders, such as glucose intolerance, hyperinsulinemia, dyslipidemia, coagulation disturbances and hypertension, is defined as the clustering of cardiovascular risk factors with insulin resistance and android obesity (Figure 1). Activators of peroxisome proliferator-activated receptors (PPARs) modulate several of these metabolic risk factors pre-disposing to atherosclerosis.

The PPAR family is constituted of PPAR α , PPAR γ and PPAR β/δ , each showing a distinct tissue distribution pattern. PPAR α is expressed preferentially in tissues where fatty acids are catabolized, whereas PPARy is highest expressed in adipose tissue where it modulates crucial aspects of adipocyte differentiation and lipid metabolism, thus impacting on glucose metabolism. PPAR β/δ is expressed at high levels in muscle and intestine, but is also present in adipose tissue and liver. All PPARs are expressed in most cell types of the vascular wall, including endothelial cells, monocyte/macrophages, smooth muscle cells and lymphocytes, as well as in atherosclerotic plaques. PPARs regulate lipid and lipoprotein metabolism and glucose homeostasis after activation by fatty acids and their metabolites or, under therapeutic conditions, by synthetic ligands, such as the hypolipidemic fibrates (low affinity PPAR α activators) and the anti-diabetic glitazones (a class of high affinity PPARy ligands). Most of the physiological functions of PPARs can be explained by their activity as transcription

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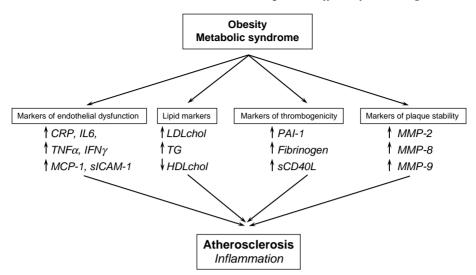


Figure 1. Biomarkers associated with features of the metabolic syndrome. The metabolic syndrome is defined as the clustering of cardiovascular risk factors pre-disposing to atherosclerosis development, such as glucose intolerance, hyperinsulinemia, dyslipidemia, coagulation disturbances and hypertension.

factors, modulating the expression of specific target genes. Upon ligand activation, PPARs regulate gene transcription by dimerizing with the Retinoid X Receptor (RXR) and binding to specific PPAR response elements (PPREs) within the regulatory regions of target genes (Chinetti et al. 2003). PPARs can also repress gene transcription in a DNA binding-independent manner by negatively interfering with other signalling pathways via protein-protein interactions and co-factor competition (Chinetti et al. 2003). Such trans-repression mechanism is likely to participate in the anti-inflammatory actions of PPARs.

PPAR agonists and metabolic effects

In humans, activation of PPAR α decreases plasma triglycerides (TG) and increases HDL cholesterol levels. The TG-lowering effects are the consequence of actions on different metabolic pathways (Staels et al. 1998a). Fibrates enhance intra-vascular triglyceride lipolysis by inducing hepatic LPL expression and activity and increasing accessibility of TG-rich lipoproteins due to reduced apoCIII content (Staels et al. 1998a). Fibrates increase FA catabolism and reduce FA synthesis, resulting in a limited TG and VLDL production in the liver. Interestingly, expression of apoAV, a recently identified determinant of plasma TG levels, is induced by fibrates in human hepatocytes (Vu-Dac et al. 2003). As a consequence of their action on lipoprotein lipolysis, these drugs also change plasma LDL composition by reducing the levels of small dense LDL, which possess lower affinity for the LDL-receptor than large LDL, thus accelerating LDL catabolism. Fibrates also influence HDL metabolism via PPARα by increasing hepatic gene transcription of apoAI and apoAII (Staels et al. 1998a). PPAR α agonists directly stimulate the first steps of the reverse cholesterol transport pathway, namely the efflux of cholesterol from peripheral cells through the induction of a number of trans-membrane receptors, such as the scavenger receptor SR-BI/CLA-1 and the ATP-binding cassette (ABC)A1 in macrophages (Chinetti et al.



2000a, 2001a). Glitazones are anti-diabetic drugs that reduce glucose and HbA1c levels in type 2 diabetic patients. In adipose tissue, PPAR γ activation induces the expression of genes controlling FA metabolism, such as LPL, CD36 and acyl-CoA synthetase, resulting in an increased FA uptake and TG storage in adipocytes, thus diverting FA away from skeletal muscle and liver and relieving their inhibitory effect on glucose metabolism and oxidation. In addition, PPARy agonists improve adipocyte insulin sensitivity and stimulate, at least in rodents, glycerol kinase activity, thus increasing the incorporation of glycerol into TG and diminishing the release of free FA by adipocytes. These effects contribute to the lowering of circulating FA levels, a biomarker of therapeutic efficacy. Analysis of the lipid profile of patients treated with glitazones demonstrated that these drugs significantly increase plasma HDL levels (Sunayama et al. 1999, Lawrence et al. 2004) and reduce small dense LDL (Hirano et al. 1998, Tack et al. 1998) (Figure 2). Since increased levels of HDL are associated with decreased risk of atherosclerosis, PPAR agonists might provide a new approach for the treatment of cardiovascular disease by increasing HDL levels and promoting reverse cholesterol transport. The influence of fibrates on cardiovascular risk has been demonstrated in a number of primary and secondary prevention as well as coronary angiography trials (LOCAT, BECAIT, VAHIT, DAIS, ...) (Chinetti et al. 2003). The beneficial effects of glitazones on cardiovascular mortality in type 2 diabetic patients is currently assessed in several prospective intervention trials, such as the RECORD, PROACTIVE and DREAM trials.

PPARs and inflammation control

The effects of PPAR agonists in the control of the inflammatory response have been observed in a variety of cell types. Activated PPARa inhibits the production of inflammatory response markers, such as endothelin-1, ICAM-1, VCAM-1 in endothelial cells and tissue factor, matrix metalloproteinase (MMP)-9 and TNF α in macrophages (reviewed in Chinetti et al. 2000b). In human aortic smooth muscle cells (SMC), PPAR α activation inhibits IL-1-stimulated IL-6 secretion and decreases IL-6 and COX2 gene transcription (Staels et al. 1998b). The role of PPARy in inflammation is more controversial due to the fact that PPAR γ -deficient mice are not viable and genetic evidence for such activity is, thus, difficult to obtain. Nevertheless, PPAR γ ligands clearly inhibit TNF α , IL-6 and IL-1 β expression in monocytes, IFN γ -induced expression of the T-cell-specific CXC-chemokines, IFN γ inducible protein 10 (IP-10), Mig, I-TAC and cell-adhesion molecules and endothelin-1 in endothelial cells (Chinetti et al. 2001b). In macrophages, PPARγ ligands repress inducible nitric oxyde synthase (iNOS) and MMP-9 expression (Chinetti et al. 2001b) and increase the production of the IL-1 receptor antagonist (Meier et al. 2002). PPARs are also expressed in immune cells (Faveeuw et al. 2000, Gosset et al. 2001) where they modulate the expression of cytokines and costimulatory molecules. In dendritic cells (DC), PPAR γ activators reduce the secretion of IL-12 (Faveeuw et al. 2000, Gosset et al. 2001), a pro-inflammatory and proatherogenic cytokine and affect the surface expression of co-stimulatory molecules, such as CD80 and CD86, and the synthesis of chemokines, including RANTES and IP-10 (Gosset et al. 2001). In T-cells, PPAR γ activators reduce the synthesis of IFN γ and TNFα (Marx et al. 2002) and decrease production of IL-2 by negatively interfering with the T-cell specific transcription factor NFAT (Yan et al. 2000).



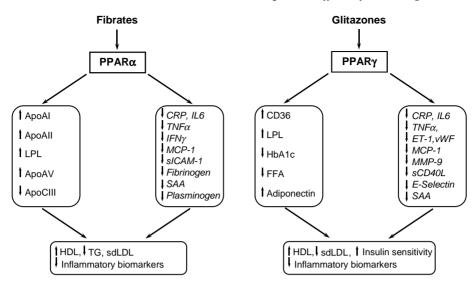


Figure 2. Effects of PPAR α and PPAR γ activators on biomarker modulation. Fibrates, through PPAR α activation, regulate the expression of genes involved in FA oxidation, TG synthesis and lipolysis and HDL metabolism. Glitazones, PPARy activators, primarily interfere with insulin signalling, FA metabolism and cytokine production. As a consequence, fibrates decrease plasma TG and small dense LDL (sdLDL) and increase HDL levels, while glitazones improve insulin sensitivity. Both drug classes reduce blood concentrations of inflammatory biomarkers, suggesting an overall beneficial effect of PPAR α and PPAR γ agonists in correcting the global risk profile pre-disposing to cardiovascular disease.

Moreover, PPARγ ligands can also influence major histocompatibility complex class (MHC) II-mediated T-cell activation by inhibiting IFNγ-induced MHC-II expression in vascular cells (Kwak et al. 2002). By contrast, the actions of PPAR α in T-cells are less well documented. In human T-cells, PPAR α activators reduce the secretion of IFN γ , TNF α and IL-2 (Marx et al. 2002). PPAR α is highly expressed in liver, where it has initially been thought to mainly regulate genes involved in fatty acid β -oxidation (Chinetti et al. 2000b). However, in primary human hepatocytes and hepatoma cells, PPARα activators suppress IL-1-induced C-reactive protein (CRP) and IL-6-induced fibrinogen expression, the major acute phase response (APR) proteins in humans (Kleemann et al. 2002), whose plasma concentrations are elevated not only in acute but also in chronic inflammatory states, such as atherosclerosis. This anti-inflammatory action of PPAR α is not restricted to these genes, but applies more generally to other APR genes, such as serum amyloid A (SAA) and fibrinogen- α and β (Gervois et al. 2001, 2004).

PPARs and biomarker modulation

Clinical trials using fibrates and glitazones also provide indications regarding the clinical efficacy of PPAR agonists in the control of inflammation. In patients with angiographically established atherosclerosis, fenofibrate treatment decreased the circulating levels of IL-6 and lowered the plasma levels of CRP and fibrinogen (Staels et al. 1998b, Gervois et al. 2004). In obese patients with atherogenic dyslipidemia, gemfibrozil administration also produced a significant reduction of CRP levels (Despres et al. 2003). In addition, fenofibrate treatment significantly reduced plasma



IFN γ and TNF α levels in patients with type IIb hyperlipoproteinemia (Madej et al. 1998). Similar effects were observed in hypertriglyceridemic patients after bezafibrate administration (Ionkers et al. 2002). Fenofibrate treatment also led to a reduction in ICAM-1, MCP-1 and α 2-macroglobulin and plasminogen plasma levels in patients with hyperlipoproteinemia (Kowalski et al. 2003, Gervois et al. 2004) (Figure 2).

Treatment of type 2 diabetic patients with rosiglitazone significantly reduced Creactive protein levels as well as white blood cell count and MMP-9 serum levels (Haffner et al. 2002). In non-diabetic coronary artery disease patients, rosiglitazone treatment resulted in a significant reduction of E-selectin, von Willebrand factor and fibrinogen as well as P-selectin levels and circulating platelet activity (Nakamura et al. 2000, Sidhu et al. 2003, 2004), while in obese patients, rosiglitazone reduced plasma concentrations of MCP-1 and TNF α , an effect already observed after 2 weeks of treatment (Mohanty et al. 2004).

In addition, in patients with coronary artery disease and type 2 diabetes, rosiglitazone significantly reduced SAA levels after only 2 weeks of treatment and decreased TNF- α levels after 6 weeks, thus suggesting an effect independent of the metabolic changes induced by the treatment (Marx et al. 2003a). Patients with type 2 diabetes exhibit elevated levels of other inflammatory markers such as sCD40L, MMP-2, MMP-3 and MMP-9, likely reflecting the increased cardiovascular risk observed in these patients (Marx et al. 2003a,b). Rosiglitazone reduced sCD40L levels as well as MMP-9 as early as 2 weeks after the initiation of treatment (Marx et al. 2003b). Interestingly, it has been reported that the maximal glucose-lowering effect of glitazones is observed only after 8-12 weeks (Raskin et al. 2000). Thus, the difference in the timing of reduction of sCD40L or MMP-9, on the one hand, and glucose, on the other hand, strongly suggests that glitazones might directly affect levels of these biomarkers, independent of their metabolic action.

Conclusions

Considerable evidence indicates that PPAR α and PPAR γ have beneficial effects in dyslipidemic and T2DM patients, as reflected by a modulation of inflammatory and cardiovascular disease biomarkers. The changes in these biomarkers reflect the role of PPARs as master regulators of lipid and lipoprotein metabolism, glucose homeostasis and the inflammatory response. Although the molecular mechanisms are not yet fully established and the complexity of the regulated systems is important, PPARs appear to interfere at different steps of atherogenesis by blocking vascular cell recruitment, by modulating foam cell formation, by interfering with the inflammatory response and by inhibiting fibrous plaque development and possibly influencing plaque stability and rupture.

Based on these findings, mainly coming from pre-clinical models as well as the first clinical reports measuring different biomarkers, fibrates, glitazones, as well as mixed PPAR α /PPAR γ ligands, can be expected to have a high potential in the treatment of inflammation-related disorders.

References

Chinetti G, Fruchart JC, Staels B. 2000b. Peroxisome proliferator-activated receptors (PPARs): nuclear receptors at the crossroads between lipid metabolism and inflammation. Inflammation Research 49:497-505.



- Chinetti G, Fruchart JC, Staels B. 2001b. Peroxisome proliferator-activated receptors (PPARs): nuclear receptors with functions in the vascular wall. Z. Kardiologia 90:125-132.
- Chinetti G, Fruchart JC, Staels B. 2003. Peroxisome proliferator-activated receptors and inflammation: from basic science to clinical applications. International Journal of Obesity Related Metabolic Disorders 27(Suppl 3):S41-S45.
- Chinetti G, Gbaguidi GF, Griglio S, Mallat Z, Antonucci H, Póulain P, Chapman J, Fruchart JC, Tedgui A, Nadib-Fruchart J, Staels B. 2000a. CLA-1/SR-BI is expressed in atherosclerotic lesion macrophages and regulated by activators of peroxisome proliferator-activated receptors. Circulation 101:2411-2417.
- Chinetti G, Lestavel S, Bocher V, Remaley A, Neve B, Torra IP, Teissier E, Minnich A, Jave M, Duverger N, et al. 2001a. PPARa and PPARy activators induce cholesterol removal from human macrophage foam cells through stimulation of the ABCA1 pathway. Nature Medicine 7:53-58.
- Despres JP, Lemieux I, Pascot A, Almeras N, Dumont N, Nadeau A, Bergeron J, Prud'Homme D. 2003. Gemfibrozil reduces plasma C-reactive protein levels in abdominally obese men with the atherogenic dyslipidemia of the metabolic syndrome. Arteriosclerosis Thrombosis and Vascular Biology 23:702-703.
- Faveeuw C, Fougeray S, Angeli V, Fontaine J, Chinetti S, Gosset P, Delerive P, Maliszewski C, Capron A, Staels B, et al. 2000. Peroxisome proliferator-activated receptor gamma activators inhibit interleukin-12 production in murine dendritic cells. FEBS Letters 486:261-266.
- Gervois P, Kleemann R, Pilon A, Percevault F, Koenig F, Staels B, Kooistra T. 2004. Global suppression of IL-6-induced acute phase response gene expression after chronic in vivo treatment with the peroxisome proliferator-activated receptor-alpha activator fenofibrate. Journal of Biological Chemistry 279:16154-
- Gervois P, Vu-Dac N, Kleemann R, Kockx M, Dubois G, Laine B, Kosykh V, Fruchart JC, Kooistra T, Staels B. 2001. Negative regulation of human fibrinogen gene expression by PPAR{alpha} agonists via inhibition of C/EBPbeta. Journal of Biological Chemistry 276:33471-33477.
- Gosset P, Charbonnier AS, Delerive P, Fontaine J, Staels B, Pastel J, Tonnel AB, Trottein F. 2001. Peroxisome proliferator-activated receptor gamma activators affect the maturation of human monocytederived dendritic cells. European Journal of Immunology 31:2857-2865.
- Haffner SM, Greenberg AS, Weston WM, Chen H, Williams K, Freed MI. 2002. Effect of rosiglitazone treatment on nontraditional markers of cardiovascular disease in patients with type 2 diabetes mellitus. Circulation 106:679-684.
- Hirano T, Yoshino G, Kazumi T. 1998. Troglitazone and small low-density lipoprotein in type 2 diabetes. Annals of Internal Medicine 129:162-163.
- Jonkers IJ, Mohrschladt MF, Westendorp RG, van der Laarse A, Smelt AH. 2002. Severe hypertriglyceridemia with insulin resistance is associated with systemic inflammation: reversal with bezafibrate therapy in a randomized controlled trial. American Journal of Medicine 112:275-280.
- Kleemann R, Gervois PP, Verschuren L, Staels B, Princen HM, Kooistra T. 2002. Fibrates down-regulate IL-1-stimulated C-reactive protein gene expression in hepatocytes by reducing nuclear p50-NF{kappa}B ~C/EBP- {beta} complex formation. Blood 101:545-551.
- Kowalski J, Okopien B, Madej A, Belowski D, Zielinski M, Kowalski J, Herman ZS. 2003. Effects of fenofibrate and simvastatin on plasma sICAM-1 and MCP-1 concentrations in patients with hyperlipoproteinemia. International Journal of Clinical Pharmacology Therapy 41:241-247.
- Kwak BR, Myit S, Mulhaupt F, Veillard N, Ruter N, Roosnek E, Mach F. 2002. PPARgamma but not PPARalpha ligands are potent repressors of major histocompatibilty complex class II induction in atheroma-associated cells. Circulation Research 90:356-362.
- Lawrence JM, Reid J, Taylor GJ, Stirling C, Reckless JP. 2004. Favorable effects of pioglitazone and metformin compared with gliclazide on lipoprotein subfractions in overweight patients with early type 2 diabetes. Diabetes Care 27:41-46.
- Madej A, Okopien B, Kowalski J, Zielinski M, Wysocki J, Szygula B, Kalina Z, Herman ZS. 1998. Effects of fenofibrate on plasma cytokine concentrations in patients with atherosclerosis and hyperlipoproteinemia IIb. International Journal of Clinical Pharmacology Therapy 36:345-349.
- Marx N, Froehlich J, Siam L, Ittner J, Wierse G, Schmidt A, Scharnagl H, Hombach V, Koenig W. 2003a. Antidiabetic PPAR gamma-activator rosiglitazone reduces MMP-9 serum levels in type 2 diabetic patients with coronary artery disease. Arteriosclerosis Thrombosis and Vascular Biology 23:283-288.
- Marx N, Imhof A, Froehlich J, Siam L, Ittner J, Wierse G, Schmidt A, Maerz W, Hombach V, Koenig W. 2003b. Effect of rosiglitazone treatment on soluble CD40L in patients with type 2 diabetes and coronary artery disease. Circulation 107:1954-1957.



- Marx N, Kehrle B, Kohlhammer K, Grub M, Koenig W, Hombach V, Libby P, Putzky J. 2002. PPAR activators as antiinflammatory mediators in human T lymphocytes: implications for atherosclerosis and transplantation-associated arteriosclerosis. Circulation Research 90:703-710.
- Meier C, Chicheportiche R, Juge-Aubry C, Dreyer M, Dayer J. 2002. Regulation of the interleukin-1 receptor antagonist in THP-1 cells by ligands of the peroxisome proliferator-activated receptor gamma. Cytokines 18:320-328
- Mohanty P, Aljada A, Ghanim H, Hofmeyer D, Tripathy D, Syed T, Al-Haddad W, Dhindsa S, Dandona P. 2004. Evidence for a potent antiinflammatory effect of rosiglitazone. Journal of Clinical Endocrinology Metabolism 89:2728-2735.
- Nakamura T, Ushiyama C, Shimada N, Hayashi K, Ebihara I, Koide H. 2000. Comparative effects of pioglitazone, glibenclamide, and voglibose on urinary endothelin-1 and albumin excretion in diabetes patients. Journal of Diabetes Complications 14:250-254.
- Raskin P, Rappaport EB, Cole ST, Yan Y, Patwardhan R, Freed MI. 2000. Rosiglitazone short-term monotherapy lowers fasting and post-prandial glucose in patients with type II diabetes. Diabetologia 43:278-284.
- Sidhu JS, Cowan D, Kaski JC. 2003. The effects of rosiglitazone, a peroxisome proliferator-activated receptor-gamma agonist, on markers of endothelial cell activation, C-reactive protein, and fibrinogen levels in non-diabetic coronary artery disease patients. Journal of the American College of Cardiology 42:1757-1763.
- Sidhu JS, Cowan D, Tooze JA, Kaski JC. 2004. Peroxisome proliferator-activated receptor-gamma agonist rosiglitazone reduces circulating platelet activity in patients without diabetes mellitus who have coronary artery disease. American Heart Journal 147:e25.
- Staels B, Dallongeville J, Auwerx J, Schoonjans K, Leitersdorf E, Fruchart J-C. 1998a. Mechanism of action of fibrates on lipid and lipoprotein metabolism. Circulation 98:2088-2093.
- Staels B, Koenig W, Habib A, Merval R, Lebret M, Torra IP, Delerive P, Fadel A, Chinetti G, Fruchart JC, et al. 1998b. Activation of human aortic smooth-muscle cells is inhibited by PPARα but not by PPARγ activators. Nature 393:790-793.
- Sunayama S, Watanabe Y, Ohmura H, Sawano M, Shimada K, Mokuno H, Daida H, Yamaguchi H. 1999. Effects of troglitazone on atherogenic lipoprotein phenotype in coronary patients with insulin resistance. Atherosclerosis 146:187-193.
- Tack CJ, Smits P, Demacker PN, Stalenhoef AF. 1998. Troglitazone decreases the proportion of small, dense LDL and increases the resistance of LDL to oxidation in obese subjects. Diabetes Care 21:796-
- Vu-Dac N, Gervois P, Jakel H, Nowak M, Bauge M, Dehondt H, Staels B, Pennacchio LA, Rubin EM, Fruchart-Nájib J, Fruchart JC. 2003. Apolipoprotein A5, a crucial determinant of plasma triglyceride levels, is highly responsive to peroxisome proliferator-activated receptor alpha activators. Journal of Biological Chemistry 278:17982-17985.
- Yan XY, Wang LH, Chen T, Hodge DR, Resau JH, DaSilva L, Farrar WL. 2000. Activation of human T lymphocytes is inhibited by peroxisome proliferator-activated receptor gamma (PPARgamma) agonists. PPARgamma co-association with transcription factor NFAT. Journal of Biological Chemistry 275:4541-4544.

